Clustering Methods for Flow Cytometry

Ultán Doherty

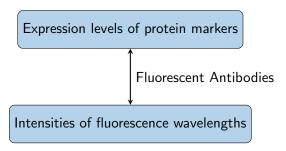
Supervisors: Arthur White & Rachel McLoughlin





Principle of Flow Cytometry

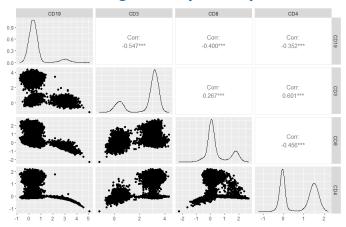
• Flow Cytometry analyses large numbers of cells to determine their expression levels of protein markers.



- Antibodies bind to specific protein markers on cells.
- Fluorochromes emit unique fluorescence wavelengths.



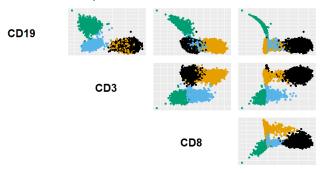
Visualising Flow Cytometry Data



 $\begin{array}{lll} \mbox{High CD19} & \Longrightarrow & \mbox{B-Cells} \\ \mbox{High CD3, High CD8} & \Longrightarrow & \mbox{Cytotoxic T-Cells} \\ \mbox{High CD3, High CD4} & \Longrightarrow & \mbox{Helper T-Cells} \end{array}$



Cell Population Identification



CD4

 $\begin{array}{lll} \mbox{High CD19} & \Longrightarrow & \mbox{B-Cells} & = \mbox{Green} \\ \mbox{High CD3, High CD8} & \Longrightarrow & \mbox{Cytotoxic T-Cells} & = \mbox{Orange} \\ \mbox{High CD3, High CD4} & \Longrightarrow & \mbox{Helper T-Cells} & = \mbox{Black} \\ \end{array}$



Sequential Manual Gating

- 1. Visualise the data in a two-dimensional plot.
- 2. Manually draw a boundary around the desired cells.
- 3. Subset the data using this boundary.
- 4. If necessary, repeat Steps 1-3 with only this subset.

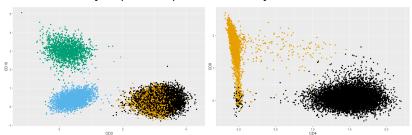


Figure: B vs T

Figure: Cytotoxic T vs Helper T



Problems with Sequential Manual Gating

- Manually drawn boundaries are biased and irreproducible.
- The sequence identifies cells with known properties.
- 2D plots do not utilise the high-dimensionality of the data.
- Subsetting can exclude interesting populations.
- Gating is time-consuming and complex for high dimensions.

Automated Cell Population Identification

- Significant research has been carried out on developing automated population identification methods.
- However, a 2020 survey revealed that 53% of laboratories never use automated flow cytometry software to identify cell populations.
 - (UK NEQAS Leucocyte Immunophenotyping survey; Cheung et al, 2021)

Difficulties with Flow Cytometry Data

- Manual gating is an unreliable benchmark for comparison.
- Populations can be skewed, heavy-tailed, and non-convex.
- Modern Flow Cytometers can measure 30-50 markers and Mass Cytometers can measure up to 100 markers.
- Each sample contains $\sim 10^5$ cells.
- Technical and biological variation exists between samples.
- Population identification must be compatible across multiple samples.

Stages of FCM Analysis

- 1a. Clustering & Meta-Clustering
 - Identify cell populations / clusters in each sample.
 - Match corresponding clusters in different samples.
- 1b. Joint Clustering
 - Identify clusters in all samples simultaneously.
 - 2. Regression
 - Predict a clinical outcome based on cluster properties.

My Research

Current Work

- Literature Review:
 - Studying existing methods and review papers.

Future Work

- Mixed Membership Model:
 - Developing a population identification method.